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# Risk Screening for Exposure to Groundwater Pollution in a Wastewater Irrigation District of the Mexico City Region

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Untreated wastewater from the Mexico City basin has been used for decades to irrigate cropland in the Mezquital Valley, State of Hidalgo, Mexico. Excess irrigation water recharges the near-surface aquifer that is used as a domestic water supply source. We assessed the groundwater quality of three key groundwater sources of domestic water by analyzing for 24 trace metals, 67 target base/neutral/acid (BNA) organic compounds, nontarget BNA organics, 23 chlorinated pesticides, 20 polychlorinated biphenyls, and nitrate, as well as microbiological contaminants—coliforms, Vibrio cholerae, and Salmonella. Study participants answered a questionnaire that estimated ingestion and dermal exposure to groundwater; 10% of the sample reported frequent diarrhea and 9% reported persistent skin irritations. Detection of V. cholerae non-01 in surface waters at all sites suggested a potential risk (surrogate indicator present) of diarrheal disease for canal and river bathers by accidental ingestion, as well as potential Vibrio contamination of near-surface groundwater and potential cholera risk, magnified by lapses in disinfection. High total coliform levels in surface water and lower levels in groundwater at all sites indicated fecal contamination and a potential risk of gastrointestinal disease in populations exposed to inadequately disinfected groundwater. Using chemical criteria, no significant risk from ingestion or dermal contact was identified at the method detection limits at any site, except from nitrate exposure: infants and young children are at risk from methemoglobinemia at all sites. Results suggest that pathogen risk interventions are a priority, whereas nitrate risk needs further characterization to determine if formal treatment is needed. The risks exist inside and outside the irrigation district. The method was highly cost-effective. Key words: groundwater, Mexico, nitrate, pathogens, risk, wastewater irrigation. Environ Health Perspect 107:553-561 (1999). [Online 3 June 1999] http://ehpnet1.niehs.nih.gov/docs/1999/107p553-561downs/abstract.html

Mexico City and the Valley of Mexico indisputably provides one of the best examples of a development model in crisis. Of the current average Mexico City wastewater flow of 45 m<sup>3</sup>/sec, approximately 75% is used, without formal treatment, to irrigate 90,000 ha in the Mezquital Valley, State of Hidalgo (1). The total human exposure implications of this water have yet to be explored. The main epidemiologic research in the region has reported significant diarrheal disease and parasitic infections in farm workers and their families (2). In the most recent review of the environmental impact of using wastewater for irrigation (arguably the largest such region in the world), the lack of attention to drinking water in the zone and potential groundwater pollution from nitrates, dissolved organic matter, and detergents was highlighted (1). Urban wastewater is transported over 80 km and distributed by canals for flood irrigation of cropland in a naturally semiarid region, recharging the local aquifer system that provides domestic water for 170,000 people. Irrigation in the Mezquital Valley was 14,000 ha in 1914-1926; 28,000 in 1950; 42,000 in 1965, and 85,000 in 1994 (3).

A recharge:discharge ratio of approximately 5:1 has been estimated (4), manifesting as a rising water table, waterlogging

of some fields, and the appearance of new springs and seeps. The region is considered among the most important in Mexico for the area it covers and its economic value from agriculture.

Mexico City Metropolitan Zone (MCMZ) wastewater is composed of a mixture of domestic, municipal, and industrial wastewater, and stormwater runoff. It receives no conventional treatment (1) and is subject to mixing and natural transformation processes during its transport from the MCMZ to the irrigation region and further biodegradation and sedimentation in a storage reservoir. Approximately 55% of the nation's industry is located in the MCMZ (paper, food, chemicals, textiles, and automotive), with approximately 43% of the wastewater in the irrigation district of industrial origin, and 57% of domestic and municipal origin (5).

The research hypothesis was that pollutant levels in groundwater represent a health risk. The research objective was to cost-effectively identify priority risks.

Study region. The study region is located 80 km north of Mexico City (Figure 1), between the Rivers Tula and Salado, in and around the Endhó Reservoir. Approximately 10,000 ha receive raw wastewater directly;

35,000 ha receive 80% wastewater + 20% fluvial reservoir water, and 25,000 ha receive naturally treated wastewater from the Requena, Endhó, Rojo Gómez, and Vicente Aguirre storage reservoirs (1).

The elevation is approximately 1,900 m above sea level, the mean temperature is 17°C, and the mean annual precipitation varies from 700 mm in the southeast to 400 mm in the north (1). Rainy season is pronounced—from June to September. Crops are alfalfa and maize (60%), oats, barley, wheat, beans, and some vegetables—chili peppers, Italian squash, and tomatoes. Cultivation of root vegetables or those consumed raw is officially prohibited, yet they are grown in some areas. Irrigation is by flooding or furrow, and rates lie between 1,500 and 2,200 mm/ha/year (1).

Population. The Mezquital population consists of municipalities of small rural villages that depend on agriculture. In the Tula Jurisdiction (irrigation district 03), 16% of the homes do not have piped water, and 45% must collect water from points outside the home. In addition, only 47% of homes have sanitation (5). Domestic water is routinely collected from groundwater wells and springs. Water is disinfected by small chlorination stations that are manually operated and maintained. The areas of focus for the project were the small village of Cerro Colorado, which is adjacent to the spring (population 110) and the area around the spring in the town of Tezontepec de Aldama (population 20,000). Because exposure is a function of socioeconomic conditions, socioeconomic characteristics of a population

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sample were gathered using a field questionnaire. Such a profile is also important for the design of appropriate interventions and risk communication strategy.

Mortality data from 1995 (6) show the top five causes of death in the State of Hidalgo (population of 2.11 million in 1995) for adults as a) heart diseases (incidence rate 61/10<sup>5</sup>); b) cirrhosis and other chronic liver diseases (46/10<sup>5</sup>); c) malignant tumors (44/10<sup>5</sup>); d) accidents (40/10<sup>5</sup>); and e) diabetes mellitus (31/10<sup>5</sup>). For infants younger than 1 year of age the top five causes were a) perinatal problems (rate 780/10<sup>5</sup>); b) congenital anomalies (257/10<sup>5</sup>); c) pneumonia and influenza (234/10<sup>5</sup>); d) gastrointestinal infectious diseases (101/10<sup>5</sup>); and e) accidents

(48/10<sup>5</sup>). All of these causes of death, except accidents, may be directly or indirectly related to environmental pollution.

Water quality assessment. Although in Mexico water quality standards are arguably the most complete in Latin America, institutional weaknesses mean they are rarely monitored and enforced (7). Growing needs for environmentally responsible wastewater reuse worldwide calls for the inclusion of a wider range of parameters that, although currently unregulated, are potentially toxic as suggested by their structure—activity characteristics: the organic fraction of known or potential toxic and mutagenic effects is the base/neutral/acid (BNA) fraction (8), such as polyaromatic hydrocarbons.

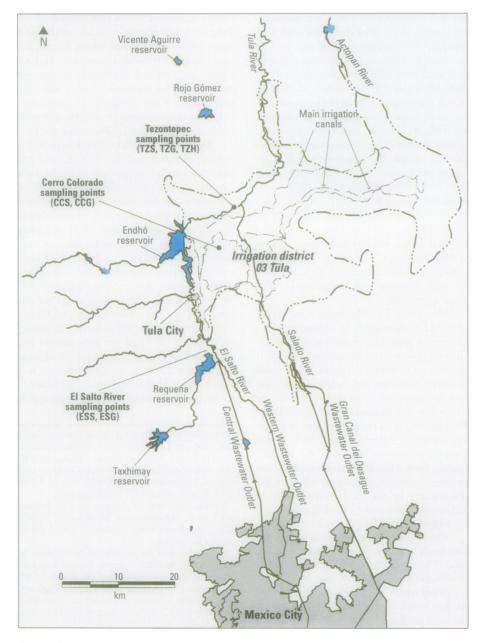


Figure 1. Study region and sampling sites.

The screening parameters consisted of 24 trace metals, 67 semivolatile BNA target organic compounds [those included in drinking water tests by U.S. Environmental Protection Agency (EPA) methods 525 and 625 (9,10)], nitrate, 23 chlorinated pesticides, and a custom 21-congener PCB suite (Table 1).

Criteria pathogens were the bacteria Vibrio cholerae and Salmonella. The V. cholerae enterotoxin produces mild to profuse diarrhea, vomiting, and rapid fluid loss. Morphologically and physiologically similar to V. cholerae 01, the non-01 vibrios produce either cholera toxin (CT) or CT-like toxin, although the diarrheal illness from their ingestion in contaminated food or water is milder (11). Only V. cholera 01 has been shown to produce cholera (12). Salmonella can cause typhoid and paratyphoid fever (13), as well as severe diarrhea and dysentery. The indicator organisms used to detect fecal pollution were total coliforms and Escherichia coli. Total coliforms include E. coli, Enterobacter, Klebsiella, and Citrobacter, and are one of the best indicators of water treatment effectiveness. Of the fecal coliforms, E. coli is the most reliable indicator because it is specifically of fecal origin (12).

#### Methods

Exposure assessment. Table 2 shows the conceptual matrix used to identify groundwater ingestion and dermal contact as local priority pathways. Exposure was estimated by applying a field questionnaire to 210 families in the Tezontepec and Cerro Colorado regions. The questionnaire asked specific questions related to ingestion and bathing in the home to quantify ingestion and dermal contact. Exposure statistics used in the United States (e.g., adult ingestion 2.0 L/day) were not assumed because climatic and cultural differences between the United States and Mexico were expected to influence values. Drinking and bathing habits were captured for two age groups: 0-15 and 16-70 years of age. All drinks and soups containing water were considered, and frequency and duration of bathing was estimated. This type of questionnaire to quantify exposure had not previously been applied in the region. Living conditions and perceptions of water problems were also captured.

Exposure to contaminants in domestic water was estimated using the pathway exposure factor (PEF) method of McKone and Daniels (14). The relationship between risk and PEF is given in Equation 1. The PEFs for ingestion and dermal contact are the exposures per unit contaminant concentration for these pathways (Equation 2). The PEFs for ingestion, PEF, and dermal

contact,  $PEF_{D}$ , are given by Equations 3 and 4, respectively. Additional infant (0–3 months) and young children ( $\leq 10$  kg) groups were used for nitrate exposure. Body weight (BW), ingestion ( $I_W$ ), and dermal contact time (T) statistics were estimated from the population questionnaire. A Monte Carlo method was used to probabilistically estimate the PEFs.

$$R = (CDI - RfD) \times p$$
 [1]

where R = individual risk (unitless probability); CDI = chronic daily intake of contaminant (mg/kg/day); RfD = reference daily intake (assumed 0 for carcinogens) (mg/kg/day); and p = contaminant toxicity or potency (mg/kg/day)<sup>-1</sup>.

$$CDI_{ij} = C_i \times PEF_j$$
 [2]

where  $CDI_{ij}$  = chronic daily intake of contaminant by exposure to medium i by pathway j (mg/kg/day);  $C_i$  = concentration of contaminant in exposure medium i (mg/L); and  $PEF_j$  = pathway exposure factor by pathway j (L/kg/day).

$$PEF_I = I_W/BW$$
 [3]

where  $I_W/BW$  = drinking water daily intake per unit body weight (L/kg/day).

$$PEF_D = T \times f_s \times SA/BW \times K_b$$
 [4]

where T = exposure duration (hr);  $f_s =$  fraction of skin surface immersed in contaminated water (unitless); SA = skin surface area (m<sup>2</sup>); BW = body weight (kg); and  $K_p =$  contaminant permeability constant across the stratum corneum (L/m<sup>2</sup>/hr).

Skin surface area (SA) was calculated from the relationship published by the International Commission on Radiological Protection (15), given in Equation 5.

$$SA = (4BW + 7)/(BW + 90)$$
 [5]

A value of  $10 \text{ L/m}^2/\text{hr}$  for  $K_p$  was used in Equation 4, following McKone and Daniels' (14) assumption that the  $K_p$  estimate for volatile organic compounds can also be used as a first-order estimate for the soluble phase of other waterborne contaminants. An estimate of  $0.6 (\pm 0.1)$  for skin fraction  $(f_s)$  immersed was used for those who bathed themselves in groundwater springs. For those who bathed using cups or buckets, dermal contact was assumed not to be a significant pathway.

Environmental sampling. Three groundwater sampling locations were chosen (Figure 1). Tezontepec is a rural town approximately 45 km from the point where

Table 1. Chemical water quality parameters.<sup>a</sup>

Name   MDL   Name   MDL   Name   Na	BNA target organic comp	ounds <sup>b</sup>	Metals	с	Chlorinated pesticides <sup>d</sup>	PCBs <sup>d</sup>
Acenaphtlylene	Name	MDL	Name	MDL		
Aniline	Acenapthene	0.06	Aluminum	14	2,4'-DDE	BZ 8
Arthracene 0.07 Beryllium 0.3 4,4'-DDT 82.44 Arboervane 0.06 Boron 51 c-BHC 82.52 Benzidine 51 Cadmium 0.99 c-Chlordane 82.68 Benzidine 0.14 Chromium 3.2 P-BHC 82.71 Benzol/filtroranthene 0.14 Chromium 3.2 P-BHC 82.71 Benzol/filtroranthene 0.15 Cobalt - cis-Chlordane 82.105 Benzol/gh/filtroranthene 0.15 Cobalt - cis-Chlordane 82.105 Benzol/gh/filtroranthene 0.15 Cobalt - cis-Chlordane 82.105 Benzol/gh/filtroranthene 0.17 Copper 1.4 c-BHC 82.118 Benzol/ghyrene 0.07 Iron 4.7 DDD+ endrin 82.126 Benzol/gh/filtroranthene 0.18 Copper 1.4 c-BHC 82.118 Benzol/ghyrene 0.07 Iron 4.7 DDD+ endrin 82.126 Benzol/gh/filtroranthene 0.18 Copper 1.4 c-BHC 82.118 Benzol/gh/filtroranthene 0.18 Copper 1.4 c-BHC 82.118 Benzol/gh/filtroranthene 0.33 Mangesium - Endosulfan uffert 82.138 Bis-12-chloroethoxylnethere 0.35 Mangenese - Endosulfan 1 82.138 Bis-12-chloroethoxylnethere 0.35 Mangenese - Endosulfan 1 82.138 Bis-12-chloroethoxylnethere 0.05 Mangenese - Endosulfan 1 82.138 Bis-12-chloroethoxylnethere 0.05 Mangenese - Endosulfan 1 82.138 Bis-12-chloroethoxylnethere 0.05 Protessium - Endosulfan 1 82.138 Bis-12-chloroethoxylnethere 0.05 Protessium - Endosulfan 1 82.138 Bis-12-chloroethoxylnethere 0.05 Protessium - Heptachlor 82.108 Bis-12-chloroethoxylnethere 0.05 Silcon - Heptachlor 7-Chloroethoxylnethere 0.05 Silcon - Heptachlor 1 Bis-12-chloroethoxylnethere 0.05 Silcon - Heptachlor						
Azobenzene						
Benzidien						
Benz/alphtroarnthene         0.06         Calcium         Aldrin         9.277           Benzo(Affluoranthene         0.14         Chromium         3.2         p.BBHC         B2 105           Benzo(Affluoranthene         0.15         Cobalt         -         cis-Chlordane         B2 105           Benzo(Aphreylene         0.37         Copper         1.4         D-BHC         B2 118           Benzo(alphrene         0.07         Iron         4.7         DDD + endrin         B2 128           Benzo(acid         4.00         Lanthanum         5.4         aldehyde           Benzyl alcohol         0.81         Lead         1.3         Dieldrin         B2 128           Bis-Q-chlorosthoxylpethane         0.33         Magnaese         Endosulfan sulfate         E2 128           Bis-Q-chlorostoxylpether         0.05         Molyddenum         —         Endosulfan 128 128           Bis-Q-chlorostoxyporpylether         0.05         Molyddenum         —         Endosulfan 128 128           Bis-Q-chlorostoxyporpylether         0.05         Molyddenum         —         Endosulfan 128 128           Bis-Q-chlorostoxyporpylether         0.02         Potassium         —         Endosulfan 28 128           Endysenzylphthene						
Benzo/ghtuoranthene         0.14         Chromium         3.2         B-BRC         BZ 101           Benzo/ghnoranthene         0.15         Cobalt         — cic-Chlordane         BZ 105           Benzo/gh/prene         0.07         Iron         4.7         DDD + endrin         BZ 128           Benzol aicid         4.00         Lead         1.3         Dieldrin         BZ 128           Benzyl alcohol         0.81         Lead         1.3         Dieldrin         BZ 128           Bis-(2-chloroethoxy)nethane         0.33         Magnesium         — Endosulfan 1         BZ 128           Bis-(2-chlorostry)lpthaleate         0.05         Manyanesum         — Endosulfan 1         BZ 128           Bis-(2-chlorostry)lpthaleate         0.05         Molyddenum         — Endosulfan 1         BZ 128           Bis-(2-chlorostry)lpthaleate         0.08         Nickel         2.2         Endrin         BZ 170           Bryberne         0.07         Silicon         — PHC         BZ 180           Bryberne         0.07         Silicon         — PHC         BZ 180           Chysene         0.07         Silicon         — PHC         BZ 180           Chyberne         0.07         Silicon         — PHC		-		0.99		
Benzo(s/h/perylene   0.15				- 22		
Benzo(a) h./perylene	• •					
Benzoialpyriane						
Benzoic acid   4.00						
Bis-12-chloroethoxylmethane Bis-12-chloroethylylether Bis-12-chloroethylylether Bis-12-chloroethylylether Bis-12-chloroethylylether Bis-12-chlorospropylether Bis-13-chlorospropylether Bis-13-chlorospr						JE 123
Bis-12-chloroethy lether   0.35	Benzyl alcohol	0.81	Lead	1.3	Dieldrin	BZ 128
Bis-   Z-ethylhexyl phthalate	Bis-(2-chloroethoxy)methane	0.33	Magnesium		Endosulfan sulfate	BZ 138
Bis-2-chloroisopropylether  4-Bromophenyl phenyl ether  Butylbenzyl phthalate  0.62  Potassium − Endrin aldehyde BZ 180  Butylbenzyl phthalate  0.30  Selenium 12  ¬PBHC BZ 195  Chrysene  0.07  Silicon − Y-Chlordane BZ 206  4-Chloroaniline  1.3  Sodium − Heptachlor DCB  4-Chloro-3-methylphenol  0.55  Strontium − Heptachlor T-Nonachl  4-Chlorophenyl-phenyl ether  0.35  Zinc  0.8  Methoxychlor  Dibenzo(a,/)anthracene  Dibenzo(a,/)anthracene  Dibenzo(a,/)anthracene  Dibenzo(a,/)anthracene  Dibenzo(a,/)anthracene  Dibenzo(a,/)anthracene  0.06  1,2-Dichlorobenzene  0.08  1,3-Dichlorobenzene  0.08  3,3-Dichlorobenzene  0.08  3,3-Dichlorobenzene  0.08  3,3-Dichlorobenzene  0.08  2,4-Dimethyl phthalate  0.06  2,4-Dimethyl phthalate  0.06  2,4-Dimethyl phthalate  0.06  1,2-Dichlorobenzene  1,2  2,6-Dinitrotoluene  1,2  2,6-Dinitrotoluene  1,2  2,6-Dinitrotoluene  0.06  Hexachlorobenzene  0.06  Hexachlorobenzene  0.06  Hexachlorobenzene  0.07  Hexachlorocyclopentadiene  1,2  2-Methyl phthalate  0.07  2-Methylphenol  0.17  2-Methylphenol  0.14  4-Methylphenol  0.14  4-Methylphenol  0.14  4-Methylphenol  0.14  4-Methylphenol  0.14  4-Methylphenol  0.14  4-Mitrophenol  0.15  2-Methylphenol  0.14  4-Mitrophenol  0.17  1.18  1.13  Nitrobenzene  0.16  2-Nitroaniline  0.17  1.14  1.15				-		
4-Bromophenyl phenyl ether         0.62         Potassium         —         Endrin aldehyde         BZ 180           Butylbenzyl phthalate         0.07         Silicon         —         γ-Chlordane         BZ 206           4-Chloro-3-methylphenol         0.55         Strontium         —         Heptachlor         DC8           4-Chloro-3-methylphenol         0.55         Strontium         —         Heptachlor         T-Nonachl           2-Chlorophenol         0.33         Vanadium         4         epoxide           4-Chloro-3-methylphenol         0.55         Zinc         0.8         Methoxychlor           Dibenzofa, Phanthracene         0.4         —         Methoxychlor         Mirex           Dibenzofaran         0.06         —         Methoxychlor         Mirex           1,2-Dichlorobenzene         0.08         —         Methoxychlor         Mirex           1,2-Dichlorobenzene         0.08         —         Methoxychlor         Mirex           1,2-Dichlorobenzene         0.08         —         Methoxychlor         Mirex           1,4-Dichlorobenzene         0.08         —         Methoxychlorobenzene         Methoxychlorobenzene         Methoxychlorobenzene         Methoxychlorobenzene         Methoxychlorobenzene <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>						
Butylbenzyl phthalate         0.30         Selenium         12         YBHC         BZ 195           Chrysene         0.07         Silicon         -         Y-Chlordane         BZ 206           4-Chloro-3-methylphenol         0.55         Strontium         -         Heptachlor         T-Nonachl           4-Chlorophenyl-phenyl ether         0.35         Zinc         0.8         Methoxychlor           Dibenzofuran         0.06         -         Nethoxychlor         Mirex           Dibenzofuran         0.06         -         -         Methoxychlor           Dibenzofuran         0.08         -         -         -           1,3-Dichlorobenzene         0.08         -         -         -         -           1,3-Dichlorobenzene         0.08         -				2.2		
Chryssen						
4-Chloro-3-methylphenol   1.3   Sodium   - Heptachlor   T-Nonachl						
4-Chloro-3-methylphenol 2-Chlorophenol 2-Chlorophenol 2-Chlorophenyl-phenyl ether 0.35 2-Chlorophenyl-phenyl ether 0.36 0.8 4-Chlorophenyl-phenyl ether 0.8 Mirex  Mirex  Dibenzofuran 0.06 0.8 1.3-Dichlorobenzene 0.08 1.3-Dichlorobenzene 0.08 3.3-Dichlorobenzene 0.08 3.3-Dichlorobenzidine 1.3 2,4-Dichlorophenol 0.61 0.62 2,4-Dimethylphenol 0.4 Dimethyl phthalate 0.06 2,4-Dimitrophenol 4.1 2,4-Dinitrotoluene 1.2 2,4-Dinitrotoluene 1.2 2,5-Dinitrotoluene 1.2 2,6-Dinitrotoluene 0.06 Hexachlorobenzene 0.06 Hexachlorobenzene 0.06 Hexachlorobenzene 0.06 Hexachlorobenzene 0.07 Hexachlorocyclopentadiene 2,4 Hexachlorocyclopentadiene 0.17 Indeno(1,2,3,4-c,dpyrene 0.37 Isophorone 0.12 2-Methylphenol 0.44 4-Methylphenol 0.49 4-Mitroaniline 0.30 1.3 Nitrobenzene 0.67 2-Nitroaniline 0.31 Nitrosodimethylamine 0.41 Nitrosodimethylamine 0.42 4-Nitrosodimethylamine 0.45 4-Nitrosodimethylamine 0.45 4-Nitrosodimethylamine 0.47 4-Nitrosodimethylamine 0.48 4-Nitrosodimethylamine 0.49 4-Nitrosodimethylamine 0.49 4-Nitrosodimethylamine 0.49 4-Nitrosodimethylamine 0.49 4-Nitrosodimethylamine 0.40 4-Nitrosodimethylamine 0.41 4-Nitrosodimethylamine 0.42 4-Nitrosodimethylamine 0.45 4-Nitrosodimethylamine 0.54 4-Nitrosodimethylamine 0.66 4-1-Nitrophenol 0.66 4-1-Nitr						
2-Chlorophenol 0.33 Vanadium 4 epoxide 4-Chlorophenyl-phenyl ether 0.35 Zinc 0.8 Methoxychlor Dibenzo(a/h)anthracene 0.4 Dibenzo(a/h)anthracene 0.04 Mirex 0.06 1.2-Dichlorobenzene 0.08 1.3-Dichlorobenzene 0.06 Dichlorophenol 0.61 Diethyl phthalate 0.06 Dichlorophenol 0.4 Dimethyl phthalate 0.06 Dichlorophenol 0.4 Dimethyl phthalate 0.06 Dichlorophenol 0.4 Dimethyl phthalate 0.06 Dichlorophenol 0.12 2.4-Dinitrotoluene 1.2 Dichlorophenol 0.06 Fluoranthene 0.06 Fluoranthene 0.06 Fluoranthene 0.06 Hexachlorobutadiene 0.09 Hexachlorophotadiene 0.79 Hexachlorocyclopentadiene 2.4 Hexachlorocyclopentadiene 2.4 Hexachlorocyclopentadiene 0.17 Indeno(1,2,3,4-c,d)pyrene 0.12 2-Methyl-4,6-dinitrophenol 0.44 4-Methylphenol 0.49 Napthalene 0.12 2-Methylphenol 0.44 4-Methylphenol 0.49 Napthalene 0.07 2-Nitroaniline 0.94 4-Nitroaniline 0.94 4-Nitroaniline 0.95 Napthalane 0.07 2-Nitroaniline 0.94 4-Nitrosodimethylamine 0.95 Napthalane 0.97 2-Nitrosodimethylamine 0.98 Nitrosodimethylamine 0.99 Nitrosodimethylamine 0.99 Nitrosodimethylamine 0.90 Nitrosodimethylamine 0.91 Nitrosodimethylamine 0.91 Nitrosodimethylamine 0.93 Pennal 0.90 Nitrosodimethylamine 0.91 Nitrosodimethylamine 0.93 Pennal 0.90 Nitrosodimethylamine 0.90 Ni				_		
4-Chlorophenyl-phenyl ether Dibenzo(a,h)ahrthacene         0.4         Zinc         0.8         Methoxychlor Dibenzola,h)ahrthacene         0.4         Mirex           Dibenzofuran         0.06         0.08         1.2-Dichlorobenzene         0.08         1.3-Dichlorobenzene         0.06         1.2-Dichlorobenzene         0.06         1.2-Dichlorobenzene         0.06         1.2-Dichlorobenzene         0.06         1.2-Dichlorobenzene         0.06         1.2-Dichlorobenzene         0.06         1.2-Dichlorobenzene         1.2-Dichlorobenzene         0.06         1.2-Dichlor				4		1-NUMBUMO
Dibenzofuran         0.4         Mirex           Dibenzofuran         0.06						
Dibenzofuran         0.06           1,2-Dichlorobenzene         0.08           1,3-Dichlorobenzene         0.08           3,3-Dichlorobenzidine         1.3           2,4-Dichlorophenol         0.61           Diethyl phthalate         0.06           2,4-Dinthyl phthalate         0.06           2,4-Dinitrophenol         4.1           2,4-Dinitrotoluene         1.2           2,6-Dinitrotoluene         1.2           2,6-Dinitrotoluene         1.2           2,6-Dinitrotoluene         0.06           Fluoranthene         0.06           Fluoranthene         0.06           Fluoranthene         0.06           Fluoranthene         0.06           Hexachlorobutadiene         0.79           Hexachlorocyclopentadiene         2.4           Hexachlorocyclopentadiene         2.4           Hexachlorocyclopentadiene         0.12           2-Methyl-4,6-dinitrophenol         2.7           2-Methyl-4,6-dinitrophenol         0.12           2-Methyl-4,6-dinitrophenol         0.44           4-Methyl-4,6-dinitrophenol         0.49           Napthalene         0.07           2-Nitrophenol         0.64           4-Nitrosodim			20	0.0		
1,3-Dichlorobenzene       0.08         1,4-Dichlorobenzene       0.08         3,3-Dichlorobenzidine       1.3         2,4-Dichlorophenol       0.61         Diethyl phthalate       0.06         2,4-Dimethylphenol       0.4         Dimethyl phthalate       0.06         Di-n-butyl phthalate       0.06         2,4-Dinitrobluene       1.2         2,4-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         1.0-n-ctyl phthalate       0.06         Huoranthene       0.06         Huoranthene       0.06         Hexachlorobuzatene       0.6         Hexachlorobutadiene       0.79         Hexachlorocyclopentadiene       0.4         Hexachlorocyclopentadiene       0.17         Hodenol 1, 2, 3, 4-c, d)pyrene       0.37         Isophorone       0.12         2-Methyl-4, 6-dinitrophenol       2.7         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       6.2	• • •					
1,4-Dichlorobenzidine       1.3         2,4-Dichlorophenol       0.61         Diethyl phthalate       0.06         2,4-Dimethylphenol       0.4         Dimethyl phthalate       0.06         2,4-Dinitrophenol       4.1         2,4-Dinitrotoluene       1.2         2,4-Dinitrotoluene       1.2         2,4-Dinitrotoluene       1.2         1,0-Di-roctyl phthalate       0.06         Huoranthene       0.06         Huoranthene       0.06         Hexachlorobenzene       0.6         Hexachlorocyclopentadiene       2.4         Hexachlorocyclopentadiene       2.4         Hexachlorocyclopentadiene       0.17         Indenol 2, 3, 4-c. plyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methyl-A G-dinitrophenol       2.7         2-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       6.2         A-Nitrosodiphenylamine       0.21         A-Nitrosodiphenylamine       0.21 <t< td=""><td>1,2-Dichlorobenzene</td><td>0.08</td><td></td><td></td><td></td><td></td></t<>	1,2-Dichlorobenzene	0.08				
3,3'-Dichlorobenzidine       1.3         2,4-Dichlorophenol       0.61         Diethyl phthalate       0.06         2,4-Dimethyl phthalate       0.06         Di-n-butyl phthalate       0.06         2,4-Dinitrotoluene       1.2         2,4-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         1:n-octyl phthalate       0.06         Fluoranthene       0.06         Fluoranthene       0.06         Hexachlorobenzene       0.6         Hexachlorobutadiene       0.79         Hexachlorocyclopentadiene       2.4         Hexachlorocyclopentadiene       0.17         Indeno(1,2,3-4-c,dpyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       0.94         4-Nitrosodinethylamine       0.83         W-Nitrosodinethylamine       0.83         W-Nitrosodinethylamine       0.21         W-Nitrosodinethylamine       0.21         W-Nitrosodinethylamine       0.21	1,3-Dichlorobenzene	0.08				
2,4-Dichlorophenol       0.61         Diethyl phthalate       0.06         2,4-Dimethylphenol       0.4         Din-butyl phthalate       0.06         2,4-Dinitrophenol       4.1         2,4-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         Di-n-octyl phthalate       0.06         Fluoranthene       0.06         Hexachlorobenzene       0.6         Hexachlorocyclopentadiene       0.79         Hexachlorocyclopentadiene       0.17         Indenol 1, 2, 3, 4-c, dipyrene       0.37         Isophorone       0.12         2-Methyl-a,6-dinitrophenol       2,7         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1,2         3-Nitroaniline       1,2         3-Nitrosodiine       0.64         4-Nitrosodinethylamine       0.64         4-Nitrosodi-p-propylamine       0.83         A-Nitrosodi-p-propylamine       0.33         Pentachlorophenol       3,1         Phenonthrene       0.06         Pyrene       0.07						
Diethyl phthalate						
2.4-Dimethyl phthalate       0.4         Dimethyl phthalate       0.06         2.4-Dinitrophenol       4.1         2.4-Dinitrotoluene       1.2         2.6-Dinitrotoluene       1.2         1.5-n-octyl phthalate       0.06         Fluoranthene       0.06         Fluorene       0.06         Hexachlorobutadiene       0.79         Hexachlorovutadiene       0.17         Indenot1,2,3-e,c,d)pyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methyl-4,6-dinitrophenol       2.7         2-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrosodinethylamine       0.83         A-Nitrosodinethylamine       0.83         A-Nitrosodinethylamine       0.21         N-Nitrosodinethylamine       0.21         N-Nitrosodinethylamine       0.21         N-Nitrosodinethylamine       0.23         Pentachlorophenol       1.5         Nenthylamine       0.06						
Dimethyl phthalate         0.06           Di-r-butyl phthalate         0.06           2,4-Dinitrophenol         4.1           2,4-Dinitrotoluene         1.2           2,6-Dinitrotoluene         1.2           Di-r-octyl phthalate         0.06           Fluoranthene         0.06           Fluoranthene         0.06           Hexachlorobenzene         0.6           Hexachlorobenzene         0.79           Hexachlorocytopentadiene         2.4           Hexachlorocytopentadiene         0.17           Indeno(1,2,3,4-c,d)pyrene         0.37           Isophorone         0.12           2-Methyl-4,6-dinitrophenol         2.7           2-Methyl-4,6-dinitrophenol         0.44           4-Methylphenol         0.49           Napthalene         0.07           2-Nitroaniline         1.2           3-Nitroaniline         1.3           Nitrobenzene         0.67           2-Nitrophenol         0.64           4-Nitrosodi-n-propylamine         0.21           A-Nitrosodi-n-propylamine         0.33           Pentachlorophenol         3.1           Phenanthrene         0.06           Pyrene         0.07						
Di-n-buryl phthalate         0.06           2,4-Dinitrophenol         4.1           2,4-Dinitrotoluene         1.2           2,6-Dinitrotoluene         1.2           Di-n-octyl phthalate         0.06           Fluoranthene         0.06           Fluorene         0.06           Hexachlorobutadiene         0.79           Hexachlorocyclopentadiene         2.4           Hexachlorocyclopentadiene         0.17           Indenol 1, 2, 3, 4-c, alpyrene         0.37           Isophorone         0.12           2-Methyl-4, 6-dinitrophenol         0.12           2-Methyl-4, 6-dinitrophenol         0.44           4-Methylphenol         0.49           Napthalene         0.07           2-Nitroaniline         1.2           3-Nitroaniline         1.3           Nitrobenzene         0.67           2-Nitrophenol         6.6           4-Nitrophenol         6.2           A-Nitrosodi-n-propylamine         0.21           A-Nitrosodi-phenol         3.1           N-Nitrosodi-phenol         3.1           Phenanthrene         0.06           Pyrene         0.07           Phenol         1.5						
2,4-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         Di-n-octyl phthalate       0.06         Fluoranthene       0.06         Hevachlorobenzene       0.6         Hexachlorocyclopentadiene       0.79         Hexachlorocyclopentadiene       0.17         Hexachlorocyclopentadiene       0.17         Hexachlorocyclopentadiene       0.12         Leyachlorocyclopentadiene       0.17         Indeno(1,2,3,4-c,d)pyrene       0.37         Isophorone       0.12         2-Methyl-4,6-dinitrophenol       2.7         2-Methyl-4,6-dinitrophenol       0.44         4-Methylphenol       0.49         4-Methylphenol       0.49         4-Nitroaniline       1.2         3-Nitroaniline       1.2         3-Nitroaniline       0.94         4-Nitrosoline       0.67         2-Nitrophenol       6.2         A-Nitrosodi-n-propylamine       0.83         A-Nitrosodi-n-propylamine       0.21         A-Nitrosodi-n-propylamine       0.31         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5						
2,4-Dinitrotoluene       1.2         2,6-Dinitrotoluene       1.2         Di-n-octyl phthalate       0.06         Fluorantene       0.06         Fluorene       0.06         Hexachlorobenzene       0.6         Hexachlorocyclopentadiene       0.79         Hexachlorocyclopentadiene       0.17         Indeno(1,2,3,4-c,d)pyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methyl-4,6-dinitrophenol       2.7         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       0.94         4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrosodimethylamine       0.83         A-Nitrosodimethylamine       0.21         A-Nitrosodin-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Di-n-octyl phthalate         0.06           Fluoranthene         0.06           Fluorene         0.06           Hexachlorobenzene         0.6           Hexachlorocyclopentadiene         0.79           Hexachloroethane         0.17           Indeno(1,2,3,4-c,d)pyrene         0.37           Isophorone         0.12           2-Methylnapthalene         0.12           2-Methyl-4,6-dinitrophenol         2.7           2-Methylphenol         0.44           4-Methylphenol         0.49           Napthalene         0.07           2-Nitroaniline         1.2           3-Nitroaniline         0.94           4-Nitroaniline         1.3           Nitrobenzene         0.67           2-Nitrophenol         0.64           4-Nitrosodimethylamine         0.83           M-Nitrosodi-n-propylamine         0.21           M-Nitrosodi-n-propylamine         0.21           M-Nitrosodi-n-propylamine         0.00           Pyrene         0.00           Phenol         1.5           2,4,5-Trichlorophenol         2.1						
Fluoranthene         0.06           Fluorene         0.06           Hexachlorobenzene         0.6           Hexachlorocyclopentadiene         0.79           Hexachlorocyclopentadiene         0.14           Hexachlorocyclopentadiene         0.17           Indeno(1,2,3,4-c,d)pyrene         0.37           Isophorone         0.12           2-Methyl-4,6-dinitrophenol         2.7           2-Methyl-4,6-dinitrophenol         0.44           4-Methylphenol         0.49           Napthalene         0.07           2-Nitroaniline         1.2           3-Nitroaniline         0.94           4-Nitroaniline         1.3           Nitrobenzene         0.67           2-Nitrophenol         6.6           4-Nitrosodiphenylamine         0.83           M-Nitrosodiphenylamine         0.21           M-Nitrosodi-n-propylamine         0.31           Pentachlorophenol         3.1           Phenonl         0.06           Pyrene         0.07           Phenol         1.5           2,4,5-Trichlorophenol         2.1						
Fluorene   0.06   Hexachlorobenzene   0.6   Hexachlorobenzene   0.6   Hexachlorobutadiene   0.79   Hexachlorocyclopentadiene   2.4   Hexachlorocyclopentadiene   0.17   Indeno(1,2,3,4- <i>c,d</i> )pyrene   0.37   Isophorone   0.12   2-Methylnapthalene   0.12   2-Methylnapthalene   0.12   2-Methylphenol   0.44   4-Methylphenol   0.49   4-Methylphenol   0.49   A-Methylphenol   0.07   2-Nitroaniline   0.94   4-Nitroaniline   1.2   3-Nitroaniline   1.3   Nitrobenzene   0.67   2-Nitrophenol   0.64   4-Nitrophenol   0.64   4-Nitrophenol   0.64   4-Nitrosodimethylamine   0.83   A-Nitrosodimethylamine   0.83   A-Nitrosodinethylamine   0.21   A-Nitrosodinethologian   0.06   Pyrene   0.07   Phenol   1.5	Di-n-octyl phthalate	0.06				
Hexachlorobenzene       0.6         Hexachlorocyclopentadiene       2.4         Hexachlorocyclopentadiene       2.4         Hexachlorochane       0.17         Indeno(1,2,3,4-c,d)pyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrosodimethylamine       0.83         A-Nitrosodiphenylamine       0.21         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenonthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Hexachlorobutadiene       0.79         Hexachlorocyclopentadiene       2.4         Hexachloroethane       0.17         Indeno(1,2,3,4-c,d)pyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methyl-4,6-dinitrophenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrophenol       6.2         M-Nitrosodimethylamine       0.83         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Hexachlorocyclopentadiene       2.4         Hexachloroethane       0.17         Indeno(1,2,3,4-c,d)pyrene       0.37         Isophorone       0.12         2-Methylnapthalene       0.12         2-Methyl-4,6-dinitrophenol       2.7         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrophenol       6.2         N-Nitrosodimethylamine       0.83         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Hexachloroethane   0.17   Indeno(1,2,3,4- <i>c,d</i> )pyrene   0.37   Isophorone   0.12   2-Methylnapthalene   0.12   2-Methyl-4,6-dinitrophenol   2.7   2-Methylphenol   0.44   4-Methylphenol   0.49   Napthalene   0.07   2-Nitroaniline   1.2   3-Nitroaniline   1.3   Nitrobenzene   0.67   2-Nitrophenol   0.64   4-Nitrophenol   0.64   4-Nitrosodimethylamine   0.83   N-Nitrosodimethylamine   0.21   N-Nitrosodi- <i>n</i> -propylamine   0.33   Pentachlorophenol   3.1   Phenanthrene   0.06   Pyrene   0.07   Phenol   1.5   2,4,5-Trichlorophenol   2.1   1.5						
Indeno(1,2,3,4- <i>c,d</i> )pyrene   0.37						
Isophorone   0.12   2-Methylnapthalene   0.12   2-Methylnapthalene   0.12   2-Methylnapthalene   0.12   2-Methylphenol   0.44   4-Methylphenol   0.49   Napthalene   0.07   2-Nitroaniline   1.2   3-Nitroaniline   1.3   3-Nitroaniline   1.3   3-Nitrobenzene   0.67   2-Nitrophenol   0.64   4-Nitrophenol   0.64   4-Nitrosodimethylamine   0.83   M-Nitrosodimethylamine   0.83   M-Nitrosodimethylamine   0.21   M-Nitrosodi-n-propylamine   0.33   Pentachlorophenol   3.1   Phenanthrene   0.06   Pyrene   0.07   Phenol   1.5   2.4,5-Trichlorophenol   2.1   2.1   3-2						
2-Methylnapthalene       0.12         2-Methyl-4,6-dinitrophenol       2.7         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       1.3         4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       6.64         4-Nitrosodimethylamine       0.83         A-Nitrosodiphenylamine       0.21         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
2-Methyl-4,6-dinitrophenol       2.7         2-Methylphenol       0.44         4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       0.94         4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       6.2         4-Nitrosodimethylamine       0.83         A-Nitrosodiphenylamine       0.21         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1	· p · · · · · · · · ·	0.12				
4-Methylphenol       0.49         Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       0.94         4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrophenol       6.2         N-Nitrosodimethylamine       0.83         N-Nitrosodi-n-propylamine       0.21         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Napthalene       0.07         2-Nitroaniline       1.2         3-Nitroaniline       0.94         4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrophenol       6.2         M-Nitrosodimethylamine       0.83         N-Nitrosodiphenylamine       0.21         M-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1		0.44				
2-Nitroaniline 1.2 3-Nitroaniline 0.94 4-Nitroaniline 1.3 Nitrobenzene 0.67 2-Nitrophenol 0.64 4-Nitrophenol 6.2 <i>I</i> -Nitrosodimethylamine 0.83 <i>I</i> -Nitrosodi- <i>n</i> -propylamine 0.33 Pentachlorophenol 3.1 Phenanthrene 0.06 Pyrene 0.07 Phenol 1.5 2,4,5-Trichlorophenol 2.1						
3-Nitroaniline       0.94         4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrosodimethylamine       0.83         N-Nitrosodiphenylamine       0.21         N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
4-Nitroaniline       1.3         Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrosodimethylamine       6.2 <i>N</i> -Nitrosodiphenylamine       0.83 <i>N</i> -Nitrosodi- <i>n</i> -propylamine       0.21 <i>N</i> -Nitrosodi- <i>n</i> -propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Nitrobenzene       0.67         2-Nitrophenol       0.64         4-Nitrophenol       6.2 <i>N</i> -Nitrosodimethylamine       0.83 <i>N</i> -Nitrosodi- <i>n</i> -propylamine       0.21 <i>N</i> -Nitrosodi- <i>n</i> -propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
2-Nitrophenol       0.64         4-Nitrophenol       6.2 <i>N</i> -Nitrosodimethylamine       0.83 <i>N</i> -Nitrosodi- <i>n</i> -propylamine       0.21 <i>N</i> -Nitrosodi- <i>n</i> -propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
4-Nitrophenol 6.2  N-Nitrosodimethylamine 0.83  N-Nitrosodi-n-propylamine 0.33 Pentachlorophenol 3.1 Phenanthrene 0.06 Pyrene 0.07 Phenol 1.5 2,4,5-Trichlorophenol 2.1						
AV-Nitrosodimethylamine       0.83         AV-Nitrosodiphenylamine       0.21         AV-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
AV-Nitrosodiphenylamine       0.21         AV-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
N-Nitrosodi-n-propylamine       0.33         Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1						
Pentachlorophenol       3.1         Phenanthrene       0.06         Pyrene       0.07         Phenol       1.5         2,4,5-Trichlorophenol       2.1	N-Nitrosodi-n-propylamine	0.33				
Pyrene         0.07           Phenol         1.5           2,4,5-Trichlorophenol         2.1	Pentachlorophenol	3.1				
Phenol 1.5 2,4,5-Trichlorophenol 2.1						
2,4,5-Trichlorophenol 2.1						
					*	
2,4,6-Trichlorophenol 0.58						

Abbreviations: BHC, benzenehexachloride (isomers); BZ, Ballschmiter-Zell classification of 209 PCB congeners; MDL, method detection limit in µg/L (ppb).

"Except nitrate [standard method 4110C (18)]. \*U.S. EPA methods 525 (drinking water) (9), 625 (wastewater) (10). \*Standard method 3120B (18). \*U.S. EPA method 608 (19).

the wastewater enters Hidalgo. It has a local spring (site TZG) derived from the infiltration of irrigation water. Cerro Colorado is the main spring in the valley, supplying domestic water to 170,000 people (Table 3). The spring (site CCG) is approximately 35 km from the wastewater entry and was the second sampling location. The third location was a natural spring derived from upland precipitation approximately 8 km from the wastewater entry. This site (ESG) near the River El Salto was the control site for near-surface groundwater quality and was a natural near-surface aguifer discharge point located outside the irrigation district. Because groundwater in the valley is either of the wastewater-derived nearsurface aquifer or the deeper (50-100 m) hydrothermal aquifer, it was important to carefully choose a control for the nearsurface aquifer.

Each sample set included one sample for metals, one for organics, and one for pathogens. Four sets of samples of groundwater were taken at each site once per month during the dry season, when dilution of contaminant concentrations was expected to be lowest (December 1996–March 1997). Samples of adjacent surface water were also taken for pathogens. Samples from a house tap at Tezontepec (site TZH) were taken only once. The sampling regime was considered optimal for the resource constraints, although some samples were lost in transit.

Microbiological testing. Water samples were collected in 200-mL plastic containers for microbiological testing at each site and at surface water sites adjacent to the groundwater sites (TZS, CCS, and ESS). Biochemical determinations of the presence/absence of V. cholerae and Salmonella were carried out at the State Public Health Laboratory (Hidalgo). Results of suspected positives were sent to the National Institute of Epidemiological Diagnosis and Reference (Santo Tomas, Mexico City, Mexico) for polymerase chain reaction confirmation. A semiportable device for quantitatively measuring total and E. coli levels, Autoanalysis Colilert (Idexx Laboratory, Inc., Westbrook, ME), was used on one sampling day. This method is also called the minimal media method (17).

Physicochemical and chemical testing. Physicochemical parameters—temperature, pH, dissolved oxygen, and conductivity—were measured in the field using portable monitors from Yellow Springs Instrument Company, Inc., Yellow Springs, Ohio. At each sampling point, amber glass bottles with Teflon (E.I. Du Pont de Nemours and Co., Wilmington, DE) caps were used to take 1 L of water for metals analysis and 1 L for organics analysis. Travel blanks consisted

Table 2. Conceptual matrix of exposure pathways.

Medium i/Pathway j	Air <i>p</i>	Air <i>g</i>	SW	GW	Soil	Sed	Biota
Inhalation	e11	e21	e31	e41	e51		-
Ingestion	e12	e22	e32	e42	e52	e62	e72ª
Dermal contact	e13	e23	e33	e43	e53	e63	_

Abbreviations: Airg, air gases; Airp, air particles; eij, possible route; eij, expected Mezquital route; eij, study focus; GW, groundwater; Sed, sediment; SW, surface water.

<sup>a</sup>In Mezquital: crops, bioconcentration of lipophylic organic compounds in milk, beef, and sheep's meat.

Table 3. Domestic water supply from Cerro Colorado spring.

Aqueduct	NS	People	Flow (L/sec)	L	Туре	Disinfection
Cerro Colorado	7	135,000	200	36	Gravity	Chlorination
C. Colorado-Huitel	23	35,000	130	7.5	Gravity	Chlorination
Total	30	170,000	330	43.5	-	_

Abbreviations: L, length (km); NS, number of settlements supplied. Data from Comisión Nacional del Agua (16).

of 1 L water purified using a Millipore complete purification system (Millipore Corp., Bedford, MA), consisting of reverse osmosis and reagent grade purification. Samples were kept in ice and shipped by express courier for next day delivery to the University of California, Los Angeles (Los Angeles, CA) laboratory.

Nitrate levels in water were determined by standard method 4110C (18). Simultaneous metals analysis of the filtered aqueous phase was carried out by standard method 3120B (18), an inductively-coupled plasma/ atomic emission (ICP/AE) method, on a Perkin-Elmer Optima 3000 DA ICP-AES Spectrometer (Perkin-Elmer, Norwalk, CT). Water samples were analyzed for EPA target drinking water BNA contaminants using an expanded list of analytes, based on EPA Methods 525 (drinking water) (9) and 625 (wastewater) (10). Method 625 was modified using a capillary column. Method detection limits are given in Table 1. Samples were analyzed for chlorinated pesticides and PCBs using methods based on EPA method 608, Pesticides and PCBs [polychlorinated biphenyls] (19).

Semivolatile BNA compound, organochlorine pesticide, and PCB analyses were carried out by concentrating the water sample using standard method 3520A (Liquid-Liquid Extraction) (18). The extraction used a 1-L sample for metals analysis and 1 L for BNA. A batch extractor was used with dichloromethane as the active agent. Samples were dried on a sodium sulfate column, concentrated to 1 mL by Kuderna-Danish evaporation, and analyzed with a Finnigan 4000 (Finnigan, San Jose, CA) gas chromatography/mass spectrometer (GC/MS). The GC column was a DB-5MS, 30 m  $\times$  0.25 mm. Six internal standards were injected into the sample at 40 ng/μL before GC/MS analysis. These standards were 1,4-dichlorobenzene-D4, napthalene-D8, acenapthene-D10, phenanthrene-D10, chrysene-D12, and perylene-D12.

Chlorinated pesticides and the selected PCB suite analyses were carried out using GC/electron capture detection (ECD) on a Varian 3500 instrument (Varian, Walnut Creek, CA) and dual columns. Method detection limits for aqueous samples of PCBs and chlorinated pesticides were 0.005–0.010 µg/L.

BNA organic analysis included distinguishing and approximately quantifying nontarget trace organics, using GC/MS profiling (20,21). Nontarget compounds were tentatively identified when possible by visually inspecting sample spectra and comparing them against closest matches from the 42,000-compound EPA/National Institutes of Health Mass Spectral Database Library (22). Because response factors were unknown for nontarget compounds, each compound's concentration was approximately quantified by comparing ratios of its spectral area to the spectral area of the closest internal standard of known concentration. Each compound has a specific scan number from the GC total ion current, which is mainly a function of molecular weight (MW) and boiling point (BP). The GC/MS output total ion chromatograms were cleaned of peaks found in travel blanks and phthalates known to be experimental artifacts. A list of target and tentatively identified nontarget compounds was compiled for each sample for data comparison.

#### **Results and Discussion**

Socioeconomic population profile. Using the questionnaire results, the sampled population of 210 families was characterized by education level, living conditions, bathing habits, drinking habits, and health signs (Table 4). The illiteracy rate was 10% in male heads of the family and 14% in mothers. Most houses had cement floors (89%) and water closets (inside 30%, outside 46%). The percentage of families without drainage was 17%; without piped water, 11%. Springs were popular bathing places (39% of

Table 4. Socioeconomic population profile.<sup>a</sup>

	Characteristic	% <i>b</i>
1.1	Age of head of family 20-30 years	22
1.2	Age of head of family 30-40 years	24
1.3	Age of head of family 60-70 years	15
2	Education	
2.1	Family head reads and writes	89
2.2 2.3	Family head neither reads nor writes	10
2.3 2.4	Mother reads and writes	83 14
2.4 2.5	Mother neither reads nor writes Family head did finish primary school	14 29
2.5 2.6	Family head did not finish primary school	37
2.0 2.7	Family head did finish secondary school	12
2. <i>1</i> 3	Living conditions	12
ა 3.1	House floor is cement	89
3.2	House floor is earth	6
3.3	House roof is tin	31
3.4	House roof is cement	68
3.5	House roof is cardboard	2
3.6	House has WC inside	30
3.7	House has WC outside	46
3.8	House has no WC	24
3.9	House has waste drainage	74
3.10	House has septic tank	8
3.1	House has no sanitation	17
3.1	House has piped water inside	27
3.1	House has piped water outside	62
3.1	House has no piped water	11
4 4.1	Bathing habits Family bathes inside with shower	25
4.2	Family bathes with small bowl	29
4.3	Family bathes in spring	39
5	Drinking habits	
5.1	Drinking water from tap	28
5.2	Drinking water from spring	65
5.3	Drinking bottled water	4
5.4	Boil water > 10 min before drinking	24
5.5	Boil water < 10 min before drinking	7
5.6	Do nothing to water before drinking	65
6	Health signs	40
6.1 6.2	Frequent diarrhea among family Persistent skin irritations	10
u.Z	reisistent skin imtations	9

WC, water closet.

 $^{a}n = 210$  families; important exposure data in bold.  $^{b}$ Percentage of sampled families with characteristic.

families) and sources of drinking water (65%). Most families did not sterilize their drinking water (65%), and only 4% used bottled water. Ten percent of families reported frequent diarrhea. Nine percent reported persistent skin irritations, which could be effects of dermal contact with wastewater. Almost all families questioned stated that the main advantage of treating wastewaters before irrigation would be "cleaner crops."

Observations of barefoot farm workers, children bathing in canals, and livestock ingesting wastewater suggest human dermal exposure to wastewater (e33, Table 2) and ingestion (e72, Table 2) of the food products of animals exposed to wastewater should be assessed—cow's and goat's milk, beef, and sheep's meat.

Groundwater pathway exposure factors. The children's mean groundwater ingestion was 1.6 ± 0.48 L/day [standard deviation

(SD)], and the adult mean was  $2.0 \pm 0.45$ L/day (Table 5). Mean dermal contact times (± SD) for children and adults were 11 ± 3.6 and 11 ± 3.3 min/day, respectively. The mean PEFs (± SDs) for ingestion for children and adults were 0.092 ± 0.076 and 0.033 ± 0.010 L/kg/day, respectively (Table 6). The mean PEFs (± SDs) for dermal contact for children and adults were  $0.035 \pm 0.017$  and  $0.024 \pm 0.010$  L/kg/day, respectively. In children, the ingestion rate  $(I_{W})$  contributed 70% of the uncertainty in the ingestion PEF and BW 30%, whereas for dermal contact 54% of the uncertainty was accounted for by uncertainty in contact time (T) and 31% by skin fraction immersed (f). For adults, the ingestion rate contributed 66% of the uncertainty in the ingestion PEF and BW 34%, whereas for dermal contact 55% of the uncertainty was accounted for by uncertainty in contact time and 34% by skin fraction immersed.

Microbiological indicators. Of the March 1997 surface water samples TZS, CCS, and ESS (adjacent to the groundwater sites TZG, CCG, and ESG), only the TZS sample was positive for the *V. cholerae* serogroup non-01. All surface water site samples taken in December 1996 were positive for V. cholerae non-01. Samples taken in December 1996 were negative for Salmonella at all sites. The March 1997 coliform counts were greater than the instrument maximum > 2,419/100 mL) for total coliforms and E. coli in all surface water sites. The March 1997 groundwater samples TZG, CCG, and ESG registered total coliforms at 770; 1,730; and 37/100 mL, respectively, and E. coli at 7, 4, and 0/100 mL, respectively. The March 1997 house sample (TZH) had values of 0 for total coliforms and E. coli.

The presence of V. cholerae-non 01 in all surface water sites is a red flag; the nearsurface groundwater supplies are potentially at risk from contamination, with a resultant potential cholera risk. Because this bacterium is almost exclusively transmitted by water (13), prevention of epidemic cholera depends on providing a safe drinking water supply that is chlorinated and free from sewage contamination; lapses in disinfection that occur during the manual changeover of chlorine tanks magnifies the risk. Coliform counts suggest a potential risk of gastrointestinal diseases at all sites, with total coliforms and E. coli counts above the Mexican standard (23) of 2/100 mL and 0/100 mL, respectively. World Health Organization (WHO) guidelines (12) for fecal coliforms in drinking water were exceeded (includes E. coli), as were the WHO guidelines (24) for wastewater reuse for the irrigation of crops likely to be eaten uncooked—limits for fecal coliforms were ≤ 1,000 per 100 mL (geometric mean). Unlike studies with wastewater reservoirs in Israel, where fecal coliforms are removed by up to five orders of magnitude after retention (25), the comparison of the site before retention in the Endhó Reservoir (Figure 1, ESG/ESS) with the site after retention (TZG/TZS) did not show this effect. This may be because the Tula River receives fecal pollution after retention, along the 20 km between the reservoir and the sampling site.

Nitrate data. Table 7 shows means and SDs of nitrate ion concentration (NO<sub>3</sub>-). Mean nitrate in groundwater ranged from 47 to 69 mg/L, whereas the health standard is 50 mg/L. The single sample at the house (TZH) was highest at 73 mg/L. ESG (control site) nitrate levels of 47 mg/L are not attributable to wastewater infiltration: manure from grazing cattle and horses, and fertilizers are likely sources. Using PEF, values from Table 6, mean CDI for the children ranged from 4.3 mg NO<sub>3</sub>-/kg/day at ESG to 6.3 mg NO<sub>3</sub>-/kg/day at CCG. The TZH value was 6.7 mg NO<sub>3</sub>-/kg/day. Corresponding values for the adult group were 36% of the child values. Assuming a risk group of young children of BW 10 kg and ingestion 1 L/day ( $PEF_I = 0.1 \text{ L/kg/day}$ ), the CDI ranges from 4.7 mg NO<sub>3</sub>-/kg/day at ESG to 6.9 mg NO<sub>3</sub>-/kg/day at CCG. For the highest risk group of infants assuming a BW of 4 kg and ingestion of 0.6 L/day (20)— $PEF_I$  on the order of 0.15 L/kg/day the CDI ranges from 7 mg NO<sub>3</sub>/kg/day at ESG to 10 mg NO<sub>3</sub>/kg/day at CCG.

In infants, the drinking water no observed adverse effect level (NOAEL) and lowest observed adverse effect level (LOAEL) for methemoglobinemia have been given in dose terms as 1.6 mg nitratenitrogen/kg/day and 1.8-3.2 mg/kg/day, respectively (26,27), equivalent to approximately 7 mg NO<sub>3</sub>-/kg/day and 8-14 mg NO<sub>3</sub>/kg/day. The NOAEL is also the oral reference dose (oral RfD) in equation 1 (uncertainty factor of 1). The infant group is at risk at all sites because CDI ≥ RfD. Using the acceptable daily intake (ADI) of 3.65 mg NO<sub>3</sub>-/kg/day, young children of BW  $\leq$  10 kg and ingestion  $\geq$  1.0 L/day are also at risk because CDI ≥ ADI. In methemoglobinemia, nitrate reduced to nitrite oxidizes hemoglobin to methemoglobin, impairing oxygen transport to tissues. Although experiments with animals suggest neither nitrate nor nitrite acts directly as a carcinogen, it may increase cancer risk in humans by endogenous formation of Nnitroso compounds whereas evidence implicating high nitrate in drinking water with other end points such as congenital malformations, cardiovascular effects, and thyroid effects are inconclusive (12).

Physicochemical data. Table 7 shows means and standard deviations of temperature at site (T), site pH, conductivity (C), dissolved oxygen (DO). Mean T was approximately 20°C, with pH ranging from 7.1 to 7.6. Conductivity ranged from 1,200 to 1,850 µmhos (TZH) and DO between 5.2 and 6.9 mg/L, with coefficients of variation at CCG and ESG of 0.45 and 0.23, respectively.

Metals, BNA organics, chlorinated pesticides, and PCBs. Mean levels of the 24 target heavy metals did not exceed U.S. Safe Drinking Water Act (29) maximum contaminant levels and maximum-level goals or Mexican standards (23) at any site; therefore, no significant risk was identified by these criteria (Table 8). No target BNA organics were detected in the samples; therefore, no significant risk was identified by these criteria at the method detection limit, whereas several nontarget BNA compounds were detected in samples, most of which could not be identified (Table 9). The number of tentatively identified compounds in the sampled groundwater ranged from 0 (ESG) to 2 (CCG), and unidentified compounds from 2 (TZG) to 9 (CCG). All tentatively identified compound levels were  $\leq 2$  $\mu$ g/L, and all detected compounds were  $\leq 6$ μg/L. Tentatively identified and unidentified compounds are of unknown but potential toxicity, so an unknown risk exists by this criterion.

Only one chlorinated pesticide was detected from the list,  $\gamma$ -chlordane in ESG, on 12 March 1997 at approximately 30 pg/L (on the order of  $10^{-11}$  g/L). There is no evidence this compound is a human carcinogen (31). Levels in water in previous studies in Hawaii (32) were on the order of 1 ng/L, and levels of magnitude  $10^{-11}$  g/L are considered low. Therefore, risk by this pollutant was considered insignificant.

Several PCB congeners were detected from the PCB list (Tables 1 and 9). The main transport medium is air, with strong sorption to suspended particles and soil. For this reason, leaching is limited and groundwater levels low (33). PCBs are classified as suspected carcinogens by the EPA on the basis of animal tests. They have a potential to cause developmental and fetotoxic effects in humans, and there is evidence that they may cause hepatoxicity (34). However, typical levels between 0.1 and 0.5 ng/L found elsewhere in drinking water represent negligible contributions to body burden as compared to food intake (35). For these reasons, the low levels detected ( $\leq 36 \text{ pg/L}$ ) were not considered a risk factor. Tables 10

Table 5. Ingestion and dermal contact statistics.

Groundwater volume ingested (L)									
Age group	BW	v1	v2	v3	v4	v5	v6	νT	T
0-15 years									
Avg	22.6	1.02	0.11	0.17	0.02	0.14	0.16	1.62	10.8
SD	11.4	0.39	0.08	0.19	0.05	0.06	0.05	0.48	3.6
n <sup>a</sup>	103 <sup>b</sup>	161	161	161	161	161	161	161	113 <sup>b</sup>
16-70 years									
Avg	63.4	1.37	0.11	0.22	0.02	0.13	0.16	2.02	10.8
SD	10.1	0.35	0.10	0.18	0.05	0.06	0.09	0.45	3.3
n <sup>c</sup>	134 <sup>b</sup>	210	210	210	210	210	210	210	150 <sup>b</sup>

Abbreviations: Avg, sample average; BW, body weight (kg); n, sample size; SD, sample standard deviation; T, daily dermal contact time (min/day) from bathing in springs; v1, water volume (liters) consumed from plain water drink; v2, flavored water drinks; v3, coffee and/or tea; v4, atole corn drink with water; v5, soup or broth; v6, stew; vT, total volume (liters). \*Number of families with children in that group. \*Sample size in age group. \*Number of families.

Table 6. Pathway exposure factor statistics.

	Children 0–1	5 years of age	Adults 16-70	years of age
	PEF <sub>I</sub>	PEF <sub>D</sub>	PEF,	$PEF_D$
Statistic				
Mean	0.092	0.035	0.033	0.024
Median	0.070	0.034	0.032	0.023
Mode	0.057	0.034	0.030	0.017
Standard deviation	0.076	0.017	0.010	0.010
Sensitivity <sup>a</sup>				
Ingestion ( $\Lambda^b$	70	_	66	4
Body weight (BW)	30	10	34	3
Bathing time (7)	_	54	_	55
Skin fraction $(f)$	_	31	-	35
Permeability $(K_p)$	_	5	_	7

Abbreviations:  $PEF_D$ , pathway exposure factor for dermal contact (L/kg/day);  $PEF_p$ , pathway exposure factor for ingestion (L/kg/day).

Percent contributions to pathway exposure factor variance. See Equations 3 and 4.

and 11 summarize chemical and microbiological health risks, respectively, identified at method detection limits.

Removal processes. The chemical contaminant levels in near-surface groundwater were lower than expected considering the short infiltration depth of a few meters for raw wastewater. However, because the sampling sites are springs, the water has also been subject to horizontal groundwater flow over distances up to several tens of kilometers. Degradation and dispersion processes, organic sorption to soil and trapping of suspended solids, and Endhó Reservoir retention (Figure 1) appear to effectively remove BNA organic contaminants. During the 80-100 km journey of wastewater from Mexico City to the Irrigation District, it is likely that metals and organics associated with suspended matter settle out, whereas the soluble fractions of metals and organics sorb to bottom sediments and soils during infiltration. Biodegradation of xenobiotics is probably active in the canals, reservoir, and soil (36). Despite the apparent efficiency of natural treatment processes, if the water table continues to rise and fields become saturated (4), the bioavailability of wastewater-borne contaminants will increase, creating different exposure conditions. This is one argument

for reducing the incoming wastewater flow from Mexico City, and this would also free up the wastewater for recycling within the Mexico City basin, where water is scarce. Using cleaner local groundwater and more efficient irrigation would reduce environmental health impacts and allow higher value crops to be grown.

Direct exposure to surface wastewater through inhalation and dermal contact, and indirectly through ingestion of milk, beef, sheep's meat, and crops may be important and should be evaluated: Dairy cattle and sheep drink directly from sewage canals because no other surface water is available.

### Conclusions and Recommendations

**Risks.** Our results suggest that pathogens are priority agents as compared to chemicals for groundwater ingestion, and they are not restricted to the wastewater irrigation district. By the coliform criterion, a potential risk of gastrointestinal disease was identified with total coliforms and *E. coli* counts above the Mexican standard (23), particularly because disinfection can be intermittent when chlorine tanks run dry and changeover is not efficient. No risk by *Salmonella* was identified. The presence of *V. cholerae* non-01 in surface waters including the river (TZS) indicates a

Table 7. Nitrate and physicochemical field data.

		NO <sub>3</sub> -	(mg/L)		Cond	uctivity	/ (µmhos	/cm)	Disse	olved o	xygen (n	ng/L)	Te	mpera	ture (°	C)		рŀ		
Site	М	SD	CV	n	M	SD	CV	n	M	SD	CV	п	M	SD	CV	n	M	SD	CV	n
Blank	ND	ND	ND	_	46	17	0.4	3	_	_	_	_	_		_	_	NM	NM	NM	_
TZG	61	2	0.0	3	1,780	66	0.0	3	6.1	0.7	0.1	3	21.4	0.1	0.0	3	7.4	0.3	0.04	3
CCG	69	1	0.0	4	1,630	550	0.3	3	5.7	2.6	0.4	3	19.7	0.4	0.0	3	7.1	0.2	0.03	3
ESG <sup>a</sup>	47	7	0.2	4	1,230	360	0.3	3	6.9	1.6	0.2	3	20.8	0.4	0.0	3	7.6	0.4	0.05	3
TZH	(73)	_	_	1	(1,850)	-		1	5.2	0.6	0.1	2	19.7	0.2	0.0	3	(7.1)	-	-	1

Abbreviations: CCG, Cerro Colorado River groundwater site; CV, coefficient of variation; ESG, El Salto River groundwater site; M, mean; n, number of samples; ND, not detected; NM, not measured; SD, standard deviation; TZG, Tezontepec de Aldama groundwater site; TZH, Tezontepec de Aldama house site. Single sample values are denoted by parentheses.

\*Control site—zero wastewater influence.

Table 8. Metals levels in groundwater (µg/L) (aqueous phase, solid phase negligible).

		Standa	rds		Bla	nks					Ground	water	levels				
Metal	Mex	MCLG	MCL	MDL	Bm	Bsd	ESGm	ESGsd	EX?	TZGm	TZGsd	EX?	TZH	EX?	CCGm	CCGsd	EX?
Aluminum	200	N/E	N/E	14	_	_	_	_	N	_	08,	N	_	N	_	ord in	N
Arsenic <sup>a</sup>	50	N/A	50	7.2	-	-	-	_	N	12	3	N	15	N	16	1	N
Barium	700	2,000	2,000	3.7	-	-	81	3	·N	78	2	N	100	N	99	9	N
Beryllium	7	0	1	0.3	-	-	-	-	N	_	-	N	_	N			N
Boron	1,000	-	1,000	51	241	324	280	0	N	753	29	N	810	N	787	72	N
Cadmium <sup>a</sup>	5	5	5	0.99	-	_	_		N			N		N			N
Calcium	_	N/E	N/E	30710	9	1	49,000	1,730	_	73,000	3,220	_	78,000	-	91,000	1,160	_
Chromium <sup>a</sup>	50 <sup>b</sup>	100	100	3.2	-	70 -			N			N		N	-	- 1	N
Cobalt	512020-03	N/E	N/E	_	-	-	_	_	_	_	_	_	_	_	_		N
Copper	2,000	1,300	ÍΤ	1.4	_	_			N	90000-		N		N			N
Iron	300	N/E	N/E	4.7			_		N	_		N	10	N			N
Lanthanum		N/E	N/E	5.4	002				93/83 <u>-</u> 387	99 <u>99992</u> 09		250200				Secretary and	100
Lead <sup>a</sup>	25	0	15	1.3	_	-	_	_	N	_	_	N		N	-		N
Magnesium		N/E	N/E	_	28	21	32,000	1,528	N	54,000	2,300	9510	30,000	orani wasa	36,000	1,160	
Manganese	150	_	200	_	130	_	_	-	N	1	_	N	_	N	2	1	N
Molybdenum	nersia ( <del>a</del> a		40	2000	1	-	3	1	N	2	1		1		2	0	
Nickel	- 11	100	100	2.2	_		_	_	N	_	0	N	ar-security	N	_	_	N
Potassium		N/E	N/E	_	_	10 TEL	29,000	1,000		27,000	1,530	-	29,000		35,000	1,160	
Selenium	-	50	50	12	_	_	_	-	N		-	N	20,000	N	-	1,100	N
Silicon		N/E	N/E	_	242	308	42,000	580		39,000	580		40,000		35,000	1,000	
Sodium	200,000	N/E	N/E	_	241	13	86,000	7,550	N	170,000	14,900	N	200,000	N	193,000	11,600	N
Strontium		N/E	N/E	689481578	2	1	730	60	05/2/20		- 1,000	_	_	_	-	- 11,000	
Vanadium		N/E	N/E	4.0	_	_	31	1	_	33	2		26		29	6	
Zinc	5,000	N/E	N/E	0.8	-	-	-	- ·	N	-	_	N	-	N	-	_	N

Abbreviations: —, analyte not detected; Bm, blank mean; Bsd, blank SD; CCGm, Cerro Colorado River groundwater site mean (n = 3); CCGsd, Cerro Colorado River groundwater site SD (n = 3); ESDm, El Salto River groundwater site mean; ESGsd, El Salto River groundwater site SD; EX?, exceeds standards?; MCL, maximum contaminant level allowed in U.S. drinking water; MCLG, maximum contaminant level goal of the U.S. Safe Drinking Water Act [data from Pontius (26)]; MDL, method detection limit (µg/L or ppb); Mex, Mexican standard (23); N, no (neither U.S. nor Mexican standards exceeded); N/A, not applicable; N/E, not established; SD, standard deviation; TT, treatment-technique dependent; TZGm, Tezontepec de Aldama groundwater site mean; TZH, Tezontepec de Aldama groundwater site SD. Analysis by standard method 31208 (18). MDL must be < standard.

<sup>a</sup>In the ATSDR/U.S. EPA top 20 hazardous substances priority list for 1997 (*30*). <sup>b</sup>Hexavalent.

potential risk of diarrheal disease for people bathing and accidentally ingesting this water. A potential risk of cholera exists because of possible near-surface groundwater contamination by *Vibrio* in surface water, a risk increased by lapses in chlorination. Ten percent of families of the sample population reported frequent diarrhea.

By the nitrate criterion, the infant and young children groups are at risk from methemoglobinemia at all sites and are not restricted to the wastewater irrigation district. This risk warrants further characterization that includes a focused epidemiologic study before the investment of scarce resources in source control and/or treatment technology can be justified. Any future treatment should be appropriate for the rural context. No risk was identified using the metals criteria at method detection

limits. By the criteria of BNA target compounds, chlorinated pesticides, and PCBs, no significant risk was identified at method detection limits. An unknown risk exists from tentatively identified and unidentified BNA compounds, although all detected compounds were at levels  $\leq 6$  µg/L. Nine percent of the sample reported persistent skin irritations, which could be effects of dermal contact with wastewater chemicals in canals and fields.

The screening method proved cost-effective in identifying priority risks in a complex pollution situation, a valuable tool in the search for health interventions that are efficient. The screening method is less costly than traditional epidemiologic studies and orients such studies, an approach particularly appropriate in less-developed countries where resources are scarce.

Potential treatment options. Microbiological risk agents require better control, with improvements sought for manual chlorination, such as low tank and irregular dose warnings, systematic tank replacement, better operator training, and/or simple automated, low-cost systems. Major plans to treat the wastewater leaving Mexico City and entering Mezquital are under review, with large-scale primary treatment and disinfection targeted at pathogen removal (7). Major treatment/disinfection options require full cost-effectiveness and impact assessment, and an understanding of the resistances of different pathogens, not merely consideration of the most studied risk from helminth eggs (1). However, even with major treatment, local pathogen risk sources would still need attention by improving basic hygiene and low-cost rural sanitation.

Table 9. BNA organics, chlorinated pesticides, and polychlorinated biphenyl data.

Parameter/site	ESG	ESG	TZG	TZG	CCG	CCG
Before/after reservoir retention? 1997 sampling date	Before 10 Jan	Before 12 Mar	After 10 Jan	After 12 Mar	10 Jan	12 Mar
BNA organic compounds						
Number of target compounds	0	0	0	0	0	0
Mass targets (%)	0	0	0	0	0	0
Number of nontarget compounds	6	3	3	3	6	11
Number tentatively identified	1	0	1	0	1	2
Number unidentified	5	3	2	3	5	9
Mass nontargets (%)	100	100	100	100	100	100
Estimated amounts (µg/L) Nontarget compounds (tentatively identified) <sup>a</sup> Cyclotrisiloxane, hexamethyl- propanoic acid, 2-methylbutyl ester 1-dodecyne	1 -		2 -	<u></u>	1 -	- 2 1
Chlorinated pesticides (pg/L) γ-Chlordane	_	30	-	_		_
Polychlorinated biphenyls (pg/L) BZ 28 BZ 66	- 26	_	- 20	- 16	- 10	11 36
T-Nonachlor	4	_	19	_	11	_
BZ 105	2	_	_	6	_	
BZ 153	_	_	36	_	_	
BZ 180	14	25	35	_	22	7
BZ 187	6	_	13	_	-	4

Abbreviations: BNA, base/neutral/acid; BZ, Ballschmiter-Zell classification of 209 PCB congeners; CCG, Cerro Colorado River groundwater site; ESG, El Salto River groundwater site; TZG, Tezontepec de Aldama groundwater site.

\*Scan numbers 496: 1.392: and 2.059, respectively.

Table 10. Summary of chemical risks (at method detection limits).

Parameter	No.	Level	Risk
Target BNA organics	0	0	No
Nontarget BNA organics	3–11	≤ 2 µg/L	?
Chlorinated pesticides	1	30 pg/L	No
Polychlorinated biphenyls	2-5	≤ 36 pg/L	No
Nitrate	_	7-10  mg/kg/day, $a > 3.65  mg/kg/day$	Yes <sup>c</sup>
Trace metals		< drinking water standard	No

Abbreviations: ?, unknown risk; ADI, acceptable daily intake; BNA, base/neutral/acid; CDI, chronic daily intake; No., number of compounds detected. Determinations of risk apply to all sampling sites Tezontepec de Aldama, Cerro Colorado River inside wastewater irrigation district, and El Salto River outside.

Infant group CDI. Young children CDI. Methemoglobinemia risk in infants (CDI ≥ reference dose) and young children (CDI > ADI).

Table 11. Summary of microbiological risks (at method detection limits).

Parameter	Presence	Risk
Vibrio cholerae	V. cholerae non-01	Potential <sup>a</sup> cholera
Salmonella	None	No
Total coliforms	37-770/100 mL	Potential <sup>a</sup> GI disease
E. coli	0-7/100 mL	Potential <sup>a</sup> GI disease

GI, gastrointestinal.

Determinations of risk apply to all sampling sites Tezontepec de Aldama, Cerro Colorado River inside wastewater irrigation district, and El Salto River outside.

ai.e., surrogate indicators of risk agent detected.

If further risk characterization proves nitrate management is needed, source identification and control should be the primary techniques. If control is impractical, low maintenance, village-scale denitrification of rural groundwater supplies would be appropriate. Such a method has been demonstrated by Silverstein et al. (37) using a novel packed tower biofilm reactor to a removal efficiency of 80–90% with

influent flow at 38 L/min, 88-110 mg/L NO<sub>3</sub>. Noyola and Morgan (38) in Mexico designed an anaerobic-anoxic-aerobic process to eliminate organic matter and nitrate in municipal and domestic wastewaters. When organic matter is low, as in Mezquital influent, the first-stage anaerobic-activated sludge reactor is excluded, which leaves two main components: an anoxic activated sludge denitrifier with upward flow, and a packed nitrifier, with recirculation between the two. For the Mezquital conditions, a second, smaller denitrifier can be placed after the first, in which methanol is dosed stochiometrically as the external carbon source. This method can achieve 95% removal of total nitrogen for flows on the order of 1.5 L/sec.

Institutional and educational interventions. Pathogen risks should be mitigated by a vigorous campaign that encourages hand-washing after defecation and before preparing and eating meals, and the boiling of drinking water from springs. Nitrate risk should be mitigated by breast-feeding infants instead of using water-based formula, and the consumption of low cost, certified purified water (an 18-L container costs \$6.00 to purchase and half a field worker's daily wage to renew-\$1.50). In developed countries, treatment plants would be installed to mitigate risk, but because physical intervention is often deemed too costly and is slow to appear in poor, rural communities of developing countries, active institutional and educational intervention to combat the nitrate and microbiological risks must precede it—and often substitute it. This should be a collaboration between public health officials (the local Sanitary Jurisdiction in Tula and the State Public Health Office in Pachuca), community leaders, and water resource agencies (local utility companies and the state-level National Water Commission, CNA-Hidalgo). Public health officials need to effect a more persistent campaign of risk communication with basic hygiene education that is culturally accepted, whereas the water resource agencies need to work more closely with researchers to design and help install sanitation that is economically and technically feasible for the rural, agricultural context of Mezquital. Environmental health problems are slowly beginning to fuel much-needed multidisciplinary collaboration at the technical level, but interinstitutional collaboration, community participation, and the transfer of knowledge into cost-effective solutions to priority problems are still the major challenges in Mexico—collective responsibilities for health professionals, engineers, and politicians to assume.

Research recommendations. Applied research should first address priority interventions for pathogen risks. Secondary topics include a fuller characterization of nitrate risk—an adequate, focused case-control epidemiologic study using a new control site exposed to low nitrate; improved surveillance (any infant death from methemoglobinemia is presently unidentified in local health records); direct and indirect human exposure to wastewater in canals and flooded fields, especially dermal contact for bathing children and barefoot farmworkers; and indirect human exposure by ingestion of crops, cow's milk, beef, and sheep's meat.

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