Clark University [Clark Digital Commons](https://commons.clarku.edu/)

[Biology](https://commons.clarku.edu/faculty_biology) [Faculty Works by Department and/or School](https://commons.clarku.edu/faculty_departments)

7-23-2020

Genome sequences of synechococcus sp. Strain MIT S9220 and cocultured cyanophage SynMITS9220M01

B. Shafer Belisle Clark University

Andres A.Avila Paz Clark University

Angelina R. Carpenter Clark University

Tayla C. Cormier Clark University

Adam J. Lewis Clark University

See next page for additional authors

Follow this and additional works at: [https://commons.clarku.edu/faculty_biology](https://commons.clarku.edu/faculty_biology?utm_source=commons.clarku.edu%2Ffaculty_biology%2F55&utm_medium=PDF&utm_campaign=PDFCoverPages)

Part of the [Biology Commons](https://network.bepress.com/hgg/discipline/41?utm_source=commons.clarku.edu%2Ffaculty_biology%2F55&utm_medium=PDF&utm_campaign=PDFCoverPages)

Repository Citation

Belisle, B. Shafer; Paz, Andres A.Avila; Carpenter, Angelina R.; Cormier, Tayla C.; Lewis, Adam J.; Menin, Linnea S.; Oliveira, Daniel R.; Song, Bukyung; Szeto, Amy; Tchantouridze, Elizabeth I.; Watson, Kayleigh A.; Yohannes, Mary T.; and Ahlgren, Nathan A., "Genome sequences of synechococcus sp. Strain MIT S9220 and cocultured cyanophage SynMITS9220M01" (2020). Biology. 55. [https://commons.clarku.edu/faculty_biology/55](https://commons.clarku.edu/faculty_biology/55?utm_source=commons.clarku.edu%2Ffaculty_biology%2F55&utm_medium=PDF&utm_campaign=PDFCoverPages)

This Article is brought to you for free and open access by the Faculty Works by Department and/or School at Clark Digital Commons. It has been accepted for inclusion in Biology by an authorized administrator of Clark Digital Commons. For more information, please contact [larobinson@clarku.edu, cstebbins@clarku.edu](mailto:larobinson@clarku.edu,%20cstebbins@clarku.edu).

Authors

B. Shafer Belisle, Andres A.Avila Paz, Angelina R. Carpenter, Tayla C. Cormier, Adam J. Lewis, Linnea S. Menin, Daniel R. Oliveira, Bukyung Song, Amy Szeto, Elizabeth I. Tchantouridze, Kayleigh A. Watson, Mary T. Yohannes, and Nathan A. Ahlgren

GENOME SEQUENCES

Genome Sequences of Synechococcus sp. Strain MIT S9220 and Cocultured Cyanophage SynMITS9220M01

Microbiology

Resource Announcements

[B. Shafer Belisle,a](https://orcid.org/0000-0002-9559-4344) Andres A. Avila Paz,a Angelina R. Carpenter,a Tayla C. Cormier,a Adam J. Lewis,a Linnea S. Menin,a Daniel R. Oliveira,a BuKyung Song,a Amy Szeto,a Elizabeth I. Tchantouridze,a Kayleigh A. Watson,a Mary T. Yohannes,a [Nathan A. Ahlgrena](https://orcid.org/0000-0001-7617-4435)

aBiology Department, Clark University, Worcester, Massachusetts, USA

AMERICAN SOCIETY FOR

MICROBIOLOGY

ABSTRACT Synechococcus bacteria are unicellular cyanobacteria that contribute significantly to global marine primary production. We report the nearly complete genome sequence of Synechococcus sp. strain MIT S9220, which lacks the nitrate utilization genes present in most marine Synechococcus genomes. Assembly also produced the complete genome sequence of a cyanophage present in the MIT S9220 culture.

arine Synechococcus bacteria are globally distributed picocyanobacteria that contribute to \sim 17% of annual net marine primary production [\(1\)](#page-4-0). The genome of Synechococcus sp. strain MIT S9220, reported here, adds to the existing six isolate genomes from the CRD1 ecotype, which exhibits physiological and genomic adaptations to its low-Fe niche [\(2,](#page-4-1) [3\)](#page-4-2). MIT S9220 was isolated from the equatorial Pacific [\(4\)](#page-4-3) and cannot grow on nitrate, in contrast to most other marine Synechococcus species [\(4,](#page-4-3) [5\)](#page-4-4).

DNA was extracted with a phenol-chloroform protocol [\(6,](#page-4-5) [7\)](#page-4-6) from a late-exponentialphase, nonaxenic culture of MIT S9220 (obtained from Gabrielle Rocap), propagated in Pro99 seawater-based medium [\(8\)](#page-4-7) at 23°C with 14:10-h light/dark illumination. Illumina sequencing (Nextera library kit; NextSeq 550) yielded 20,222,448 paired 150-bp reads, and Oxford Nanopore MinIon sequencing (library kit SQK-RAD004; R9.4.1 flow cell) produced 53,387 reads (60 Mb; N_{50} , 1,727 bp). The Illumina reads were quality filtered with Trimmomatic v. 0.38 [\(9\)](#page-4-8) with the following settings: ILLUMINACLIP:TruSeq3-PE-2.fa:2:30:10, LEADING:10, TRAILING:10, SLIDINGWINDOW:4:15, and MINLEN:50. Assembly with Unicycler v. 0.4.7 [\(10\)](#page-4-9) with default settings using both Illumina and Nanopore reads yielded 13 contigs, 12 of which belong to MIT S9220 (2,406,645 total bp; N_{50} , 1,568,415 bp; GC content, 56.4%; 300 \times mean coverage) based on high similarity to other CRD1 contigs (blastn; E value, <1e-84; identity, ≥88%). The MIT S9220 genome was annotated with the Prokaryotic Genome Annotation Pipeline (PGAP) v. 4.11 [\(11\)](#page-4-10), and it contains 2,575 protein-coding genes, 42 tRNAs, and 2 rRNA operons. We estimate that the genome is 99% complete based on it possessing 1,026 homologs (reciprocal best hits; blastp; E value, <1e-10) of 1,033 single-copy core genes shared among Synechococcus isolate genomes [\(3,](#page-4-2) [12\)](#page-4-11). Like other CRD1 genomes, MIT S9220 possesses a larger repertoire of Fe-related genes than most other Synechococcus genomes [\(3\)](#page-4-2).

MIT S9220 lacks the nitrate utilization genes found in most other marine Synechococcus species [\(Fig. 1\)](#page-3-0) [\(13\)](#page-4-12). A similar, parallel loss [\(3\)](#page-4-2) occurs in Synechococcus sp. RS9917 (ecotype VIII) [\(Fig. 1\)](#page-3-0) [\(14\)](#page-4-13). MIT S9220 provides a valuable resource for further exploring the evolution of nitrogen utilization in these important phytoplankton, especially given similar loss and occasional reacquisition of these genes in closely related Prochlorococcus species [\(15,](#page-4-14) [16\)](#page-4-15).

The remaining circular, 190,237-bp contig (38.8% GC content, 150 \times coverage),

Citation Belisle BS, Avila Paz AA, Carpenter AR, Cormier TC, Lewis AJ, Menin LS, Oliveira DR, Song B, Szeto A, Tchantouridze EI, Watson KA, Yohannes MT, Ahlgren NA. 2020. Genome sequences of Synechococcus sp. strain MIT S9220 and cocultured cyanophage SynMITS9220M01. Microbiol Resour Announc 9:e00481-20. [https://doi.org/10.1128/MRA](https://doi.org/10.1128/MRA.00481-20) [.00481-20.](https://doi.org/10.1128/MRA.00481-20)

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2020 Belisle et al. This is an openaccess article distributed under the terms of the [Creative Commons Attribution 4.0](https://creativecommons.org/licenses/by/4.0/) [International license.](https://creativecommons.org/licenses/by/4.0/)

Address correspondence to Nathan A. Ahlgren, [nahlgren@clarku.edu.](mailto:nahlgren@clarku.edu)

Received 28 May 2020 **Accepted** 4 July 2020 **Published** 23 July 2020

FIG 1 Regions in the MIT S9220 and RS9917 genomes (names in purple) that exhibit the loss of nitrate utilization genes (purple trapezoids) found in all other marine Synechococcus genomes sequenced to date. The nitrate operon includes genes encoding nitrate reductase (narB), a nitrate transporter (napA), and genes for synthesis of Mo cofactor required for nitrate reduction (moaA to moaE, mobA, and moeA). For reference, homologous regions are shown for the genomes of WH7803 and WH8102, which both have the nitrate utilization operon and either lack or possess, respectively, urea utilization genes (urtA to urtE and ureA to ureG) near the nitrate operon. Also labeled are the nitrate reduction (nirA) and transport (focA) genes and several conserved genes that flank the nitrate operon (pyrG, queE, cobA, and cynG).

named SynMITS9220M01, is a Caudovirales T4-like myovirus based on high similarity to other cyanophage (top blastn results against the NCBI nonredundant [nr] database; E value, 0). SynMITS9220M01 could be an extrachromosomal phage, but it more likely represents an unintentionally cocultured phage obtained from unautoclaved, 0.2 - μ mfiltered Sargasso Sea water used for culture medium for several months during its propagation. SynMITS9220M01 is most similar to cyanophage S-SM2 [\(17\)](#page-4-16) (average nucleotide identity [ANI], 72.3%). SynMITS9220M01 was annotated with Prokka v. 1.13.3 with default settings except for the option "–protein" used with a database of 325 marine cyanophage genomes. SynMITS9220M01 has 10 tRNAs and 239 predicted protein-coding genes, including the following host-acquired genes: phosphate-related genes (phoH, pstS) [\(17](#page-4-16)[–](#page-4-17)[20\)](#page-4-18), photosynthesis genes (psbA, psbD, petE) [\(18,](#page-4-19) [21,](#page-4-20) [22\)](#page-4-21), and others, such as talC, mazG [\(23](#page-4-22)[–](#page-4-23)[25\)](#page-4-24), and genes involved in heptose-related lipopolysaccharide synthesis (D,D-heptose 7-phosphate kinase, phosphoheptose isomerase, and ADP-L-glycero-D-mannoheptose-6-epimerase) [\(26\)](#page-4-25). SynMITS9220M01 adds to the growing database of 300 marine cyanophage genomes [\(27\)](#page-4-26) and to our knowledge of cyanophage pangenomic diversity.

Data availability. Synechococcus sp. strain MIT S9220 is available from the Roscoff Culture Collection [\(http://roscoff-culture-collection.org/;](http://roscoff-culture-collection.org/) strain RCC2571) or from Nathan A. Ahlgren upon request. Sequence data are available at NCBI under BioProject number [PRJNA623799,](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA623799) including the raw data (accession numbers [SRX8340787](https://www.ncbi.nlm.nih.gov/sra/SRX8340787) and [SRX8330445\)](https://www.ncbi.nlm.nih.gov/sra/SRX8330445) and the assembled genomes (accession numbers [JABBNJ000000000](https://www.ncbi.nlm.nih.gov/nuccore/JABBNJ000000000) and [MT408532\)](https://www.ncbi.nlm.nih.gov/nuccore/MT408532).

ACKNOWLEDGMENTS

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sector.

The genomes reported were generated by the collaborative efforts of graduate and undergraduate students in Clark University's The Genome Project course (BIOL209).

REFERENCES

- 1. Flombaum P, Gallegos JL, Gordillo RA, Rincón J, Zabala LL, Jiao N, Karl DM, Li WKW, Lomas MW, Veneziano D, Vera CS, Vrugt JA, Martiny AC. 2013. Present and future global distributions of the marine cyanobacteria Prochlorococcus and Synechococcus. Proc Natl Acad Sci U S A 110:9824 –9829. [https://doi.org/10.1073/pnas.1307701110.](https://doi.org/10.1073/pnas.1307701110)
- 2. Sohm JA, Ahlgren NA, Thomson ZJ, Williams C, Moffett JW, Saito MA, Webb EA, Rocap G. 2016. Co-occurring Synechococcus ecotypes occupy four major oceanic regimes defined by temperature, macronutrients and iron. ISME J 10:333–345. [https://doi.org/10.1038/ismej.2015.115.](https://doi.org/10.1038/ismej.2015.115)
- 3. Ahlgren NA, Belisle BS, Lee MD. 2020. Genomic mosaicism underlies the adaptation of marine Synechococcus ecotypes to distinct oceanic iron niches. Environ Microbiol 22:1801–1815. [https://doi.org/10.1111/1462](https://doi.org/10.1111/1462-2920.14893) [-2920.14893.](https://doi.org/10.1111/1462-2920.14893)
- 4. Moore LR, Post AF, Rocap G, Chisholm SW. 2002. Utilization of different nitrogen sources by the marine cyanobacteria Prochlorococcus and Synechococcus. Limnol Oceanogr 47:989 –996. [https://doi.org/10.4319/lo](https://doi.org/10.4319/lo.2002.47.4.0989) [.2002.47.4.0989.](https://doi.org/10.4319/lo.2002.47.4.0989)
- 5. Fuller NJ, Marie D, Partensky F, Vaulot D, Post AF, Scanlan DJ. 2003. Clade-specific 16S ribosomal DNA oligonucleotides reveal the predominance of a single marine Synechococcus clade throughout a stratified water column in the Red Sea. Appl Environ Microbiol 69:2430 –2443. [https://doi.org/10.1128/aem.69.5.2430-2443.2003.](https://doi.org/10.1128/aem.69.5.2430-2443.2003)
- 6. Green MR, Sambrook J. 2017. Isolation of high-molecular-weight DNA using organic solvent. Cold Spring Harb Protoc 2017:pdb.prot093450. [https://doi.org/10.1101/pdb.prot093450.](https://doi.org/10.1101/pdb.prot093450)
- 7. Wurch L, Giannone RJ, Belisle BS, Swift C, Utturkar S, Hettich RL, Reysenbach AL, Podar M. 2016. Genomics-informed isolation and characterization of a symbiotic Nanoarchaeota system from a terrestrial geothermal environment. Nat Commun 7:12115. [https://doi.org/10.1038/](https://doi.org/10.1038/ncomms12115) [ncomms12115.](https://doi.org/10.1038/ncomms12115)
- 8. Moore LR, Coe A, Zinser ER, Saito MA, Sullivan MB, Lindell D, Frois-Moniz K, Waterbury J, Chisholm SW. 2007. Culturing the marine cyanobacterium Prochlorococcus. Limnol Oceanogr Methods 5:353-362. [https://doi](https://doi.org/10.4319/lom.2007.5.353) [.org/10.4319/lom.2007.5.353.](https://doi.org/10.4319/lom.2007.5.353)
- 9. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114 –2120. [https://doi.org/10](https://doi.org/10.1093/bioinformatics/btu170) [.1093/bioinformatics/btu170.](https://doi.org/10.1093/bioinformatics/btu170)
- 10. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. [https://doi.org/10.1371/journal.pcbi.1005595.](https://doi.org/10.1371/journal.pcbi.1005595)
- 11. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614 – 6624. [https://doi.org/10.1093/nar/gkw569.](https://doi.org/10.1093/nar/gkw569)
- 12. Lee MD, Ahlgren NA, Kling JD, Walworth NG, Rocap G, Saito MA, Hutchins DA, Webb EA. 2019. Marine Synechococcus isolates representing globally abundant genomic lineages demonstrate a unique evolutionary path of genome reduction without a decrease in GC content. Environ Microbiol 21:1677–1686. [https://doi.org/10.1111/1462-2920.14552.](https://doi.org/10.1111/1462-2920.14552)
- 13. Moreno-Vivian C, Cabello P, Martinez-Luque M, Blasco R, Castillo F. 1999. Prokaryotic nitrate reduction: molecular properties and functional distinction among bacterial nitrate reductases. J Bacteriol 181:6573– 6584. [https://doi.org/10.1128/JB.181.21.6573-6584.1999.](https://doi.org/10.1128/JB.181.21.6573-6584.1999)
- 14. Dufresne A, Ostrowski M, Scanlan D, Garczarek L, Mazard S, Palenik B, Paulsen I, de Marsac N, Wincker P, Dossat C, Ferriera S, Johnson J, Post A, Hess W, Partensky F. 2008. Unraveling the genomic mosaic of a

ubiquitous genus of marine cyanobacteria. Genome Biol 9:R90. [https://](https://doi.org/10.1186/gb-2008-9-5-r90) [doi.org/10.1186/gb-2008-9-5-r90.](https://doi.org/10.1186/gb-2008-9-5-r90)

- 15. Berube PM, Biller SJ, Kent AG, Berta-Thompson JW, Roggensack SE, Roache-Johnson KH, Ackerman M, Moore LR, Meisel JD, Sher D, Thompson LR, Campbell L, Martiny AC, Chisholm SW. 2015. Physiology and evolution of nitrate acquisition in Prochlorococcus. ISME J 9:1195–1207. [https://doi.org/10.1038/ismej.2014.211.](https://doi.org/10.1038/ismej.2014.211)
- 16. Berube PM, Rasmussen A, Braakman R, Stepanauskas R, Chisholm SW. 2019. Emergence of trait variability through the lens of nitrogen assimilation in Prochlorococcus. Elife 8:e41043. [https://doi.org/10.7554/eLife](https://doi.org/10.7554/eLife.41043) [.41043.](https://doi.org/10.7554/eLife.41043)
- 17. Sullivan MB, Huang KH, Ignacio-Espinoza JC, Berlin AM, Kelly L, Weigele PR, DeFrancesco AS, Kern SE, Thompson LR, Young S, Yandava C, Fu R, Krastins B, Chase M, Sarracino D, Osburne MS, Henn MR, Chisholm SW. 2010. Genomic analysis of oceanic cyanobacterial myoviruses compared with T4-like myoviruses from diverse hosts and environments. Environ Microbiol 12:3035–3056. [https://doi.org/10.1111/j.1462-2920.2010.02280.x.](https://doi.org/10.1111/j.1462-2920.2010.02280.x)
- 18. Mann NH, Clokie MR, Millard A, Cook A, Wilson WH, Wheatley PJ, Letarov A, Krisch HM. 2005. The genome of S-PM2, a "photosynthetic" T4-type bacteriophage that infects marine Synechococcus strains. J Bacteriol 187:3188 –3200. [https://doi.org/10.1128/JB.187.9.3188-3200.2005.](https://doi.org/10.1128/JB.187.9.3188-3200.2005)
- 19. Sullivan MB, Coleman ML, Weigele P, Rohwer F, Chisholm SW. 2005. Three Prochlorococcus cyanophage genomes: signature features and ecological interpretations. PLoS Biol 3:e144. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pbio.0030144) [journal.pbio.0030144.](https://doi.org/10.1371/journal.pbio.0030144)
- 20. Kelly L, Ding H, Huang KH, Osburne MS, Chisholm SW. 2013. Genetic diversity in cultured and wild marine cyanomyoviruses reveals phosphorus stress as a strong selective agent. ISME J 7:1827–1841. [https://doi](https://doi.org/10.1038/ismej.2013.58) [.org/10.1038/ismej.2013.58.](https://doi.org/10.1038/ismej.2013.58)
- 21. Lindell D, Sullivan MB, Johnson ZI, Tolonen AC, Rohwer F, Chisholm SW. 2004. Transfer of photosynthesis genes to and from Prochlorococcus viruses. Proc Natl Acad Sci U S A 101:11013-11018. [https://doi.org/10](https://doi.org/10.1073/pnas.0401526101) [.1073/pnas.0401526101.](https://doi.org/10.1073/pnas.0401526101)
- 22. Puxty RJ, Perez-Sepulveda B, Rihtman B, Evans DJ, Millard AD, Scanlan DJ. 2015. Spontaneous deletion of an "ORFanage" region facilitates host adaptation in a "photosynthetic" cyanophage. PLoS One 10:e0132642. [https://doi.org/10.1371/journal.pone.0132642.](https://doi.org/10.1371/journal.pone.0132642)
- 23. Thompson LR, Zeng Q, Kelly L, Huang KH, Singer AU, Stubbe J, Chisholm SW. 2011. Phage auxiliary metabolic genes and the redirection of cyanobacterial host carbon metabolism. Proc Natl Acad Sci U S A 108: E757–E764. [https://doi.org/10.1073/pnas.1102164108.](https://doi.org/10.1073/pnas.1102164108)
- 24. Bryan MJ, Burroughs NJ, Spence EM, Clokie MR, Mann NH, Bryan SJ. 2008. Evidence for the intense exchange of MazG in marine cyanophages by horizontal gene transfer. PLoS One 3:e2048. [https://doi](https://doi.org/10.1371/journal.pone.0002048) [.org/10.1371/journal.pone.0002048.](https://doi.org/10.1371/journal.pone.0002048)
- 25. Clokie MR, Mann NH. 2006. Marine cyanophages and light. Environ Microbiol 8:2074 –2082. [https://doi.org/10.1111/j.1462-2920.2006.01171.x.](https://doi.org/10.1111/j.1462-2920.2006.01171.x)
- 26. Kneidinger B, Graninger M, Puchberger M, Kosma P, Messner P. 2001. Biosynthesis of nucleotide-activated D-glycero-D-manno-heptose. J Biol Chem 276:20935–20944. [https://doi.org/10.1074/jbc.M100378200.](https://doi.org/10.1074/jbc.M100378200)
- 27. Gregory AC, Solonenko SA, Ignacio-Espinoza JC, LaButti K, Copeland A, Sudek S, Maitland A, Chittick L, Dos Santos F, Weitz JS, Worden AZ, Woyke T, Sullivan MB. 2016. Genomic differentiation among wild cyanophages despite widespread horizontal gene transfer. BMC Genomics 17:930. [https://doi.org/10.1186/s12864-016-3286-x.](https://doi.org/10.1186/s12864-016-3286-x)